



Review Article



The Power of Nanovaccines in Immunotherapy of Melanoma, Lung, Breast, and Colon Cancers: A Comprehensive Review

Seyedeh Ghazaleh Angaji¹ , Mohammad Amin Salim² , Alireza Azizi³ , Negin Amiri⁴ , Saeede Rastakhiz⁵ , Negar Jahani⁶ , Behnaz Akhlaghi⁷ , and Parsa Ebrahimi Tirtashi^{8,*}

¹ Faculty of Pharmacy, Alborz University of Medical Sciences, Karaj, Iran

² Faculty of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran

³ Faculty of Pharmacy, Yeditepe University, Istanbul, Turkey

⁴ Department of Pharmaceutics, School of Pharmacy and Novel Drug Delivery Systems Research Centre, Isfahan University of Medical Sciences, Isfahan, Iran

⁵ Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

⁶ Student Research Committee, Faculty of Pharmacy, Mashhad University of Medical Sciences, Mashhad, Iran

⁷ Department of Quality Control of Drug Products, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran

⁸ Faculty of Pharmacy, Azad University of Medical Sciences, Tehran, Iran

* **Corresponding author:** Parsa Ebrahimi Tirtashi, Faculty of Pharmacy, Azad University of Medical Sciences, Tehran, Iran. Email: parsaebrahimi1379@gmail.com

ARTICLE INFO

Article History:

Received: 14/10/2023

Revised: 14/11/2023

Accepted: 26/11/2023

Published: 25/12/2023



Keywords:

Breast cancer

Cancer immunotherapy

Colon cancer

Lung cancer

Melanoma

Nanovaccines

ABSTRACT

Scientists are exploring new approaches to overcome cancer, and nanovaccines have emerged as one of the most promising tools in the fight against cancer. This review aimed to provide a thorough overview of nanovaccines as potential cancer immunotherapy agents by describing their mechanism of action and potential therapeutic implications. The growing incidence of cancer underscores the urgent need for comprehensive strategies focusing on prevention, early detection, and innovative treatment modalities to control and mitigate the impact of this widespread disease effectively. It is important to note that nanovaccines are a cutting-edge platform with a wide range of applications in immunotherapy for colon, breast, lung, melanoma, and ovarian cancers. Nanoscale formulations of tumor-specific antigens and adjuvants can initiate an efficient and targeted immune response. Research on nanovaccines involving melanoma has shown that they can trigger potent anti-tumor immune responses, which permit prolonged survival and tumor regression. Furthermore, nanovaccines have been effective in treating breast cancer since they can modulate the tumor microenvironment and stimulate the presence of cytotoxic T cells within the tumor. The nanovaccines strategy has enhanced the immune system's recognition of tumor antigens, resulting in tumor cell destruction and effective immune recognition. There have also been studies that have utilized nanovaccines to modify the immune response of tumor cells to immune checkpoint inhibitors, thereby improving the synergistic outcomes of colon cancer treatment. Besides improving the immune response to malignancies, nanovaccines represent a transformative approach to cancer immunotherapy. The presence of compelling results across various cancer types suggests that nanovaccines are a powerful tool in cancer treatment despite further research required to optimize their design and validate their clinical applicability.

1. Introduction

The incidence of various malignant tumors has been steadily increasing, making cancer the leading cause of death in recent years^{1,2}. In this regard, immunotherapy has

played an important role in cancer treatment, standing as the fourth treatment pathway following surgery, chemotherapy, and radiotherapy as the fourth major

► *Cite this paper as:* Angaji SGh, Salim MA, Azizi A, Amiri N, Rastakhiz S, Jahani N, Akhlaghi B, Ebrahimi Tirtashi P. The Power of Nanovaccines in Immunotherapy of Melanoma, Lung, Breast, and Colon Cancers: A Comprehensive Review. Research in Biotechnology and Environmental Science. 2023; 2(4): 55-64. DOI: 10.58803/rbes.v2i4.21



The Author(s). Published by Rovedar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

treatment pathway³. *In-vitro* experiments have shown that certain parasites, including the protozoans *Toxoplasma gondii*, *Trypanosoma cruzi*, and the helminths *Trichinella spiralis* and *Echinococcus granulosus* represent anti-cancer activities⁴⁻⁷. Nanovaccines are a promising strategy due to their increased lowest immunotoxicity, antigen stability, flexibility of the physical characteristics of nanomaterials, and sustained release. Nano vaccines surpass regular vaccines in efficiency owing to their controllable and flexible physicochemical properties⁸. Identifying and eliminating neoplasm cells is how the immune system fights tumors. Free cancer cells are eliminated by immunotherapy, thus preventing tumor recurrence and metastasis⁹.

An antigen-based vaccine is administered to provoke an immune response against a particular pathogen⁹. Hundreds of millions of lives have been saved since the advent of vaccines in the last century. Since peptide subunit vaccines are safe and easy to produce, they are an excellent option for cancer immunotherapy¹¹. However, their immune responses are suboptimal, primarily because they are not co-delivered to lymph nodes (LNs) where B- and T-cells coordinate immune responses¹². Peptide-based vaccines are frequently combined with adjuvants and administered through delivery systems to enhance their immunogenicity¹³. Although depot-forming immunoadjuvants, like incomplete Freund's adjuvant (IFA), enhance Ag immunogenicity by releasing it sustainably from a depot, evidence suggests that persisting Ag depots may result in T-cell sequestration, exhaustion, and dysfunction. This, in turn, may prevent T cells from infiltrating tumors from vaccination sites. To date, no treatment is compelling for cancer¹⁴.

Among the many therapies available, chemotherapy is widely used throughout the world¹⁵. Patients suffer from side effects from chemotherapy, which cause inconvenience for them¹⁶. Therapeutically potent drugs are destroying the rapidly multiplying cells. In addition to killing the malignant cells, chemotherapy induces several side effects¹⁷. Due to its oppressive side effects, chemotherapy is often avoided by patients. The non-targeted distribution of chemotherapeutic drugs in the body as 'free drugs' limits the drug concentration at impaired organ sites due to non-targeted distribution throughout the body. These obstacles might hamper degenerative diseases, which are closely related to their clinical failure. Chemotherapy could be improved with site-specific targeted drug delivery. Nanotechnology involves chemistry, physics, engineering, biology, and medicine^{15,17}. It is a significant technique for treating degenerative diseases, detecting tumors early, acting specifically on them, reducing multidrug resistance, reducing toxicity, discovering cancer biomarkers, and developing novel treatments. Drug molecules have been successfully delivered to targeted sites/cells using nanocarriers¹⁸. Furthermore, nanotechnology and nanocarrier-based drug delivery systems offer improved healing efficacy and decrease undesirable side effects associated with

conventional drugs, introducing new classes of therapeutics and persuading the development of biologically active molecular new entities previously considered undevelopable due to pharmaceutically suboptimal properties^{19,20}. Nanotechnology has introduced peptide-based subunit called nanovaccines, where antigens and adjuvants are co-delivered. This innovative approach aims to optimize peptide-based subunit vaccines by leveraging various nanocarriers designed to efficiently deliver epitopes and enhance the effectiveness of nanovaccines.

In addition to offering the capability to modify their surface for functionalization and charge delivery, nanocarriers are also helpful in encapsulating loaded charges and being able to be accepted by antigen-presenting cells (APCs)²¹. Nanovaccines that deliver antigens/adjuvants promote maturation and APC activation, resulting in active T lymphocytes that attack cancer cells²². As a result of engineering approaches to the co-delivery of antigens/adjuvants to immune cells, robust immune responses can be stimulated, which in turn increases the speed and duration of immune responses, intensifies weak antigen immunogenicity, and modulates Ag-antibody responses. The first part of this study will focus on cancer vaccines and adjuvants in immunotherapy for cancer. The second part will discuss the strategies for designing and developing nanovaccines, emphasizing the interactions between epitopes and adjuvants.

2. Methods and materials

A thorough literature search, encompassing PubMed, Scopus, and Web of Science databases, was conducted between 2015 and 2023 to identify studies on nanovaccines in cancer immunotherapy. Inclusion criteria comprised research articles and reviews in English, focusing on nanovaccines development for breast cancer, colon cancer, lung cancer, melanoma, and relevant cancer types. Exclusion criteria included studies not directly related to nanovaccines or not in English. Data on nanovaccines formulations, mechanisms of action, and therapeutic outcomes were extracted, and the quality of the derived articles was checked. Due to the qualitative nature of this review, there was no need to perform statistical analysis. The review was structured to provide insights into mechanisms and implications of nanovaccines across various cancers, acknowledging potential limitations, such as variability in study designs and evolving nature of the field.

3. Cancer immunotherapy

As cancer therapy advances, immune checkpoint-based therapies, including immune checkpoint blockade, are ideal strategies for making fantastic progress²². Nevertheless, these immunotherapies are only effective for a small percentage of patients. The selection of patients is, therefore, an essential factor in preventing the harmful

effects of treatment and avoiding unnecessary costs. An early indication of response and clinical benefit will be required by identifying and validating reliable surrogate biomarkers. There is evidence that immunotherapy is particularly effective in highly mutagenized tumors²³. Mutational load contributes to the clinical response to immunotherapy via neo-antigen-specific responses²⁴. It has been reported by two independent groups that anti-CTLA-4 treatment correlated with mutational frequency in melanoma tumors²⁵.

Further, patients with colon and non-small cell lung cancer (NSCLC) treated with anti-PD1 inhibitors exhibited higher numbers of mutations, including mutations in DNA repair pathways²⁶. The anti-PD1 treatment has practical clinical effects on some kidney cancer patients with low mutational frequencies, and this is not true for all tumor types²⁷. There is a correlation between lymphocyte infiltrates and improved survival for many types of cancer²⁸. The expression of PD-L1 on tumor cells can be considered a valuable biomarker to determine which patients would benefit from immune checkpoint blockade monotherapy in addition to the use of PD-L1 on tumor cells. It has been found that some patients with high levels of PD-L1 are not responsive to programmed cell death protein 1 (PD1) pathway blockade, in contrast to those with PD-L1-negative tumors²⁹. Therefore, the lack of PD-L1 expression around the tumor microenvironment cannot reliably exclude patients from PD1 pathway blockade treatment. Therefore, PD-L1 is not an optimal biomarker for patient selection³⁰. Murine models have indicated that the combination of immune checkpoint inhibitors can overcome the exhausted phenotype in several tumors by characterizing TILs, including overexpression of exhaustion markers such as PD1, LAG3, and TIM3³¹. Different immunotherapeutic combinations may also respond differently based on the patient's immune system³². A system that considers the immune milieu in addition to PD-L1 status and lymphocyte profile will be essential to guide therapeutic combinations. Moreover, immunohistochemistry and genetic profiling of the tumor microenvironment could be combined to improve biomarker algorithms by classifying cancers according to their immunoevasion strategies³³.

4. Immunotherapy by vaccines

The development of vaccines and the creation of vaccines against various diseases have significantly impacted global health since the invention of vaccines³⁴. Aside from its role in protecting the body from parasites, viruses, and bacteria, the immune system also protects against cancer through its relentless fight against infections³⁵. The Food and Drug Administration (FDA) has approved five cancer vaccines up to this point; they protect against virus-induced cancers, specifically cervical cancer caused by human papillomavirus (HPV) and liver cancer caused by hepatitis B virus (HBV)³⁶. Additionally, therapeutic vaccines targeting existing

tumors are being studied along with prophylactic vaccines.

Therapeutic cancer vaccines aim to regulate tumor growth, initiate tumor regression, and eradicate existing tumors by provoking acquired immune responses in patients against specific tumor antigens³⁷. High-quality antigens must be delivered to dendritic cells (DCs) as a significant component of effective therapeutic cancer vaccination³⁸. These DCs must be optimally activated to generate persistent and robust responses from CD4+ helper T cells and cytotoxic T lymphocytes, infiltration and recruitment into tumor microenvironments, and maintenance and durability of their effects³⁹. However, despite the difficulties in producing vaccines and their low clinical response rates, they remain attractive due to their specificity, safety, tolerability, and ability to provide long-lasting memory responses among the several strategies in cancer immunotherapy⁴⁰. The vaccine platform employed in cancer vaccines could be one reason for low response rates. A cancer vaccine can be made from various platforms, including whole cells, nucleic acids (DNA and RNA), bacterial and viral vectors, and proteins/peptides⁴¹. Constructing DNA vaccines by cloning plasmids that encode tumor antigens to elicit and enhance CD8+ and CD4+ T-cell responses is possible. Apoptosis or exosomes release transgenes in transfected cells after DNA plasmids are administered to them⁴². To amplify adaptive immune responses, DCs endocytose released transgenes and present them with Major Histocompatibility Complex I (MHC) and MHC II to CD4+ and CD8+ T-cells, respectively. A phase III trial evaluates VGX-3100 against high-grade cervical intraepithelial neoplasia by combining it with electroporation against HPV-16/HPV-18 E6 and E7 coding DNA vaccines. The phase 2b trial results showed that 49.5 % of treated women regressed their lesions, while 30.6% of placebo patients wholly regressed⁴³.

There are differences between RNA-based vaccines and DNA-based vaccines as far as their susceptibility to degradation by RNases is concerned. However, modified nucleosides incorporated and applied can protect RNA-based vaccines⁴³. DNA vaccines require transcription through the nuclear membrane barrier, whereas RNA vaccines do not⁴⁴. One clinical trial examines the tolerability, effectiveness, and safety of a personalized mRNA vaccine for advanced esophageal and non-small cell lung cancer. In a recent study, A—using an *in-vitro* transcribed (IVT) mRNA vaccine encoding four antigens failed to demonstrate satisfactory anti-tumor activity despite remarkable innovations in the design and development of mRNA-based vaccines⁴⁵. The recognition of receptors on immune cells makes vector-based vaccines an attractive delivery system for antigens and immunomodulatory molecules, resulting in the development of both acquired and innate immune systems⁴⁶. Despite this, antiviral immune responses limit the efficacy of repeated vaccinations by limiting vaccination frequencies. To address this limitation, various strategies have been employed, such as heterologous

prime-boost administration and surface coating strategies⁴⁷. RNA-based vaccines in cancer treatment are currently being evaluated through many clinical trials. Three major categories of tumor antigens can be used to develop anti-tumor vaccines, such as Tumor-Specific Antigens (TSA), which are specifically expressed in cancer cells, and Tumor-Associated Antigens (TAAs) that are expressed both on average and in cancer cells but are overexpressed on cancer cells; neoantigens that are derived from tumor cells and have unique epitopes of self-antigens. Numerous clinical trials have examined peptide-based vaccines, yet no substantiated evidence has emerged to support their remarkable advantages. In the realm of cancer vaccines, the utilization of short peptides, comprising fewer than 15 amino acids, has proven unsuccessful in eliciting robust immune responses.

Moreover, short peptides cannot fully activate CD4+ helper T-cells, which are necessary for optimal CTL activation. Several methods for overriding these limitations include combining immunostimulatory molecules with peptides, combining other adjuvants with peptides, and synthesizing long peptides (SLPs) of multiple lengths^{48,49}. Due to the tedious and expensive process of producing neoantigen-based peptide vaccines, advances in databasing, software predictions, and sequencing have shed light on overcoming the difficulties^{50,51}.

5. Nanovaccines

There are several advantages to nanoparticle (NP)-based vaccines compared to subunit Ag-based vaccines. Antigens are either encapsulated within the NPs or decorated on the surface of the NPs⁵². NPs could elicit robust immune responses due to long-term bioavailability and sustained release of Ag in contrast to soluble Ag-based vaccines. Also, NPs allow antigens to be taken up and processed by APCs, resulting in the maturation of APCs⁵³. This further promotes Ag cross-presentation by MHC class I to CD8+ T-cells, which determines innate and adaptive immune responses through the production of cytokines²².

Furthermore, NPs serve as delivery mechanisms for cytokines and activate innate immune receptors to produce them. In addition to their composition, nanoparticles' physical characteristics, such as charge, pore size, and charge density, may affect their immunogenicity⁵⁴. Nanoparticles made of mesoporous silica with extensive pores have the potential to enhance DC activation and Ag presentation, leading to the stimulation of immune responses and the inhibition of tumor growth. The utilization of nanoparticles with spacious pores holds promise in clinical applications, as it facilitates the introduction of minimal quantities of antigens and immunostimulatory molecules into the body⁵⁵.

Additionally, the size of NPs has a significant influence on immune responses. A study has shown that NPs with

smaller sizes (20–30 nm) are endocytosed by DCs, whereas macrophages phagocytose those with larger sizes (less than 0.5 μ m)⁵⁶. Additionally, the size of nanoparticles influences the efficacy of nanovaccines by determining the leakiness of tumor vasculatures and their pore sizes. The microparticle forms of alum-based adjuvants promote Th2-biased immune responses, while their nanoparticulated forms can stimulate Th1-biased immune responses to develop cancer vaccines⁵⁷. Additionally, the surface charge of nanoparticles plays an essential function in the internalization and trafficking of antigens. The electrostatic interaction between cationic NPs and the oppositely-charged cellular membrane makes them more efficient at being taken up by the APCs. The positive charges also aid in cationic NP escape from the lysosome. The blood circulation time of slightly negatively charged nanoparticles is longer than that of cationic nanoparticles of the same size⁵⁸.

Another factor that stimulates immune solid responses is the morphology of nanovaccines. According to Gong et al., the tumor antigens are trapped in endosomes and have a low immunogenicity⁵⁹. The vaccine is configured with proton-driven nanotransformers capable of altering the morphology of the endosome in an acidic environment. In response to the morphological change in nanovaccines from nanosphere to nanosheet, the endosomal membrane was disrupted, which allowed the antigenic peptide to be released into the cytoplasm. As a result, inflammation pathways were activated, and tumor growth was suppressed in HPV-induced cancer and mice models of melanoma⁵⁶.

Compared with rod-like and spherical particles, cylindrical hydrogel nanoparticles can present antigens to APCs for lasting periods, which results in robust immune responses. In a study, softer NPs showed increased circulation and targeting compared to harder ones. Also, the softer nanoparticles significantly reduced tumor cell uptake, endothelial cell uptake, and immune cell⁶⁰.

6. Mechanisms of action of nanovaccines

6.1 Antigen presentation

Nanovaccines enhance cancer immunotherapy by optimizing antigen presentation. Some nanovaccines carry antigens specific for tumors or antigenic peptides that mimic antigens present in tumors⁶¹. Antigen-presenting cells, such as dendritic cells, present the antigens to antigen-presenting cells. Nanoparticles facilitate antigen uptake and processing by APCs through various mechanisms, including receptor-mediated endocytosis. These APCs present antigens on their cell surfaces through their MHC molecules once they have internalized the antigens. By presenting antigens efficiently, T cells, specifically CD8+ cytotoxic T cells, are primed to recognize and attack cancer cells with the same antigen. A successful immune response against a tumor depends on this process⁶².

6.2 Immune cell activation

Besides delivering antigens, nanovaccines are designed to contain immune-stimulating molecules and adjuvants⁶³. These components potently activate the immune system. The adjuvant may consist of a toll-like receptor agonist, cytokines, or other immunomodulators. They activate APCs and subsequent immune responses in nanovaccines when co-delivered with antigens⁶⁴. Antigen-presenting cells release many pro-inflammatory cytokines and costimulatory molecules during activation that stimulate the activation and proliferation of effector T cells, especially CD8+ cytotoxic T cells⁶⁵. When these activated T cells are transported to the tumor site, they can kill tumor cells.

6.3 Tumor microenvironment modulation

Tumor microenvironments are often immunosuppressive environments, hindering immune responses to cancer⁶⁶. Nanovaccines may be essential in modulating the tumor microenvironment to encourage an immune response against the tumor. It is possible to manufacture nanovaccines that contain agents capable of targeting

immune suppressive cells within the TME, such as myeloid-derived suppressor cells (MDSCs) and regulatory T cells (Tregs)⁶⁷. Nanovaccines can improve anti-tumor responses by reducing the presence or activity of immunosuppressive cells. Additionally, some nanovaccines may have components that promote better immune surveillance and tumor cell recognition by infiltrating immune cells into tumors⁶⁸.

6.4 Role of adjuvants and immune-stimulating molecules

Nanovaccines comprise immune-stimulating molecules and adjuvants⁶⁹. The purpose of adjuvants is to enhance vaccine immunogenicity. Activating pattern recognition receptors (PRRs) on APCs triggers signaling pathways that induce stimulation and pro-inflammatory responses⁶⁵. For example, toll-like receptor agonists, such as lipopolysaccharides or CpG oligonucleotides, can activate TLRs on APCs, enhancing cytokine production and antigen presentation. The immune system can also be stimulated by encapsulating cytokines such as tumor necrosis factor-alpha (TNF-A) and interleukin-12 (IL-12)⁷⁰ (Figure 1).

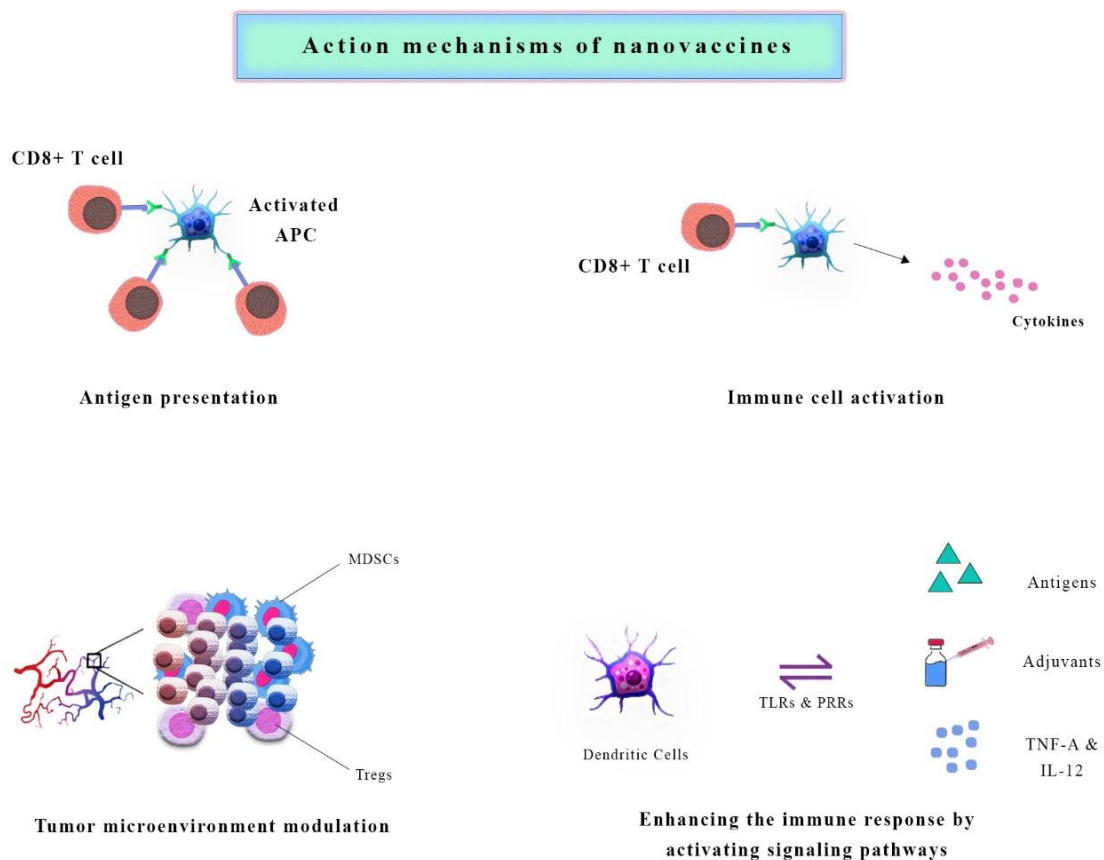


Figure 1. This figure shows the key elements of nanovaccines, highlighting the role of adjuvants and immune-stimulating molecules. The nanovaccines consist of antigens and adjuvants. The adjuvants activate pattern recognition receptors (PRRs) on antigen-presenting cells (APCs) such as dendritic cells (DCs). This interaction triggers signaling pathways leading to stimulation and pro-inflammatory responses. Toll-like receptor (TLR) agonists, like lipopolysaccharides and CpG oligonucleotides, activate TLRs on APCs, enhancing cytokine production and antigen presentation. Additionally, cytokines such as tumor necrosis factor-alpha (TNF-A) and interleukin-12 (IL-12) are encapsulated within the nanovaccines to stimulate the immune system. These components collectively enhance vaccine immunogenicity.

7. Clinical trials

Clinical trials using nanomaterials for cancer treatments mainly focus on camptothecin, paclitaxel, and monoclonal antibodies that block immune checkpoints (ICBs)⁷¹. In clinical trials, the results of nanomaterials used in cancer vaccines have been less promising than those of traditional cancer drugs. Nanomaterials are often complex in design and need modification, resulting in difficulty in mass production and quality control⁷². The clinical trial of a nanoparticle vaccine against Epstein-Barr virus (EBV) gp350 is noteworthy⁷³. Animal studies have shown that the vaccine induces high neutralizing antibodies in nonhuman primates and mice⁷⁴. Many cancers are associated with EBV, including non-Hodgkin's lymphoma, Hodgkin's lymphoma gastric cancer, and nasopharyngeal cancer⁷⁵. Therefore, it was included in clinical trials related to cancer vaccines as a preventive vaccine. The effects of treating advanced solid tumors with Poly(lactic-co-glycolic) Acid (PLGA) nanoparticles containing antigens and adjuvants are also being evaluated in phase I clinical trials⁷⁶. These vaccine components include antigens derived from the NY-ESO-1 cancer-testis antigen peptide and the alpha-galactosylceramide-derived iNKT cell activator IMM60. Phase II clinical trials (NCT00199901) have previously combined NY-ESO-1 with ISCOMATRIX® adjuvant for treating melanoma, a cellular target found in many cancers⁷⁷. Although the survival period for most patients had not improved, significant antibody responses had been induced among them, which is encouraging for future research⁷⁸. A novel cancer vaccine based on NY-ESO-1-derived peptides and IMM60 was developed using polymer PLGA nanoparticles.

Clinical trials are anticipated to demonstrate a better immune response when vaccines are nanosized, and new adjuvants are added. The mRNA-2752 and mRNA-2416 were encapsulated in lipid nanoparticles in these two clinical trials to be administered intratumorally alone or combined with Durvalumab to treat relapsed or refractory solid tumors and lymphomas^{79,80}. A recent clinical study was conducted to determine whether corresponding mRNA vaccines would be safe and tolerable for cancer patients⁸¹. Vaccines for cancer are most likely to be used as adjuncts to surgery or chemotherapy in the clinic. Preparing cancer nanovaccines formulations is time-consuming and labor-intensive in most preclinical studies. Because cancer patients' disease progression is rapid, lead times for treatment should not be too long, and cancer vaccines prepared by overly complex methods are rarely clinically useful. The immunogenicity and toxicity of vaccines with mature preparation processes are disappointing in clinical settings. In order to optimize drug delivery, FDA-approved biocompatible materials should be used, and processes should be simplified. Additionally, laboratory animal models differ significantly from the human immune system, which makes preclinical findings

challenging to translate into human studies⁸². Despite the current negative clinical results of cancer nanovaccines, as the preparation of complex nanoparticles is simplified and matured, more and more effective nanovaccines will enter clinical trials and prove effective.

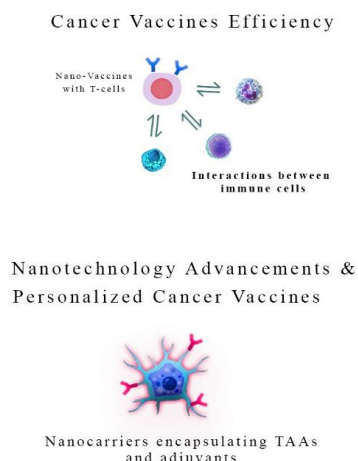
8. Future perspectives

Many successful cases of immunotherapy in cancer treatment have occurred recently, including immune checkpoint blockade and chimeric antigen receptor cells. Cancer vaccines induce an anti-tumor immune response effectively and safely as part of immunotherapy. As an adjuvant therapy, cancer vaccines enhance the efficacy of other immunotherapies. A more significant potential for cancer immunotherapy can be realized by developing nanotechnology⁸³. Various bio-based and synthetic nanocarriers have been discussed as potential cancer vaccine carriers. Through interactions between TAAs and adjuvants, they are encapsulated in nanocarriers and subsequently targeted to APCs, significantly increasing their immunogenicity⁸⁴. Vaccine components are controlled in their release by selecting the appropriate carrier or modifying it chemically. Preclinical and clinical studies are already attracting much attention in preclinical and clinical settings. Cancer nanovaccines have shown promising results in recent years as a possible therapeutic approach, but translational research still faces many challenges⁸⁵. Several challenges need to be tackled. For instance, in preclinical investigations, the majority of vaccines demonstrated positive outcomes but encountered setbacks in clinical trials. This discrepancy could be attributed to variations in the immune systems between laboratory animal models and humans, although the use of humanized mice can help mitigate some of these differences.

Developing new experimental animal tumor models that mimic human disease mechanisms is critical for a successful clinical translation of cancer vaccines⁸⁶. The selection of tumor animal models must also be based on the specific scientific question since there is a large variety of them. As cancer treatments become increasingly personalized, it is imperative to establish xenograft models derived from patients. For cancer nanovaccines to exert their efficacy, nanoparticles should be resistant to non-specific protein binding, non-toxic, and rapidly cleared by the body. Mass production and inter-batch quality controls are also essential for vaccines to reach clinical practice and be widely commercialized. Recent years have increased interest in neoantigen-based cancer vaccines due to tumor heterogeneity and individual differences⁸⁷. A critical step in developing personalized vaccines is optimizing the identification of tumor-specific antigens. Despite this, it may still be challenging to sequence antigens efficiently and accurately with the available technology. To achieve more precise sequencing and identification of novel antigens, it is imperative to integrate biological sciences with artificial intelligence and computer simulation

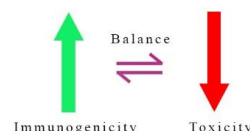
Nanovaccines in Cancer Immunotherapy

Benefits & Advancements



Challenges & Considerations

Balancing Immunogenicity and Toxicity



Lab-to-Clinic Hurdles & Biodistribution

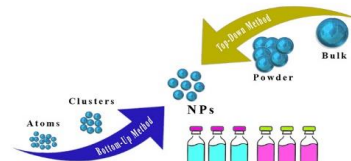


Figure 2. The benefits and advancements: Depicting the effectiveness of cancer vaccines, nanotechnology contributions, and the concept of personalized treatment. Challenges and considerations: Emphasizing the need to balance immunogenicity with potential toxicity and the hurdles in translating laboratory research into clinical applications.

techniques. This fusion of disciplines enhances the leveraging of cross-disciplinary advantages for optimal results⁸⁸. Some nanomaterials are intrinsically immunogenic, allowing them to act as carriers and adjuvants simultaneously. Thus, it is also essential to balance immunogenicity with toxicity when choosing materials. Nanoparticles can also cause problems with immune response and cytotoxicity. The use of advanced emerging technologies in cancer treatments is promising. It is possible to explore a wide range of possibilities. These emerging technologies may produce unexpected results when combined with chemotherapy or immunotherapy⁸⁹ (Figure 2)

9. Conclusions

Preclinical studies have shown that cancer vaccines are integral to immunotherapy and can be administered as single drugs or combined with other agents. Nanomaterial-based cancer vaccines are at the beginning of their development, and they still face obstacles in clinical settings. Nanovaccines have the potential to emerge as the next generation of cancer immunotherapy, evolving alongside advancements in diverse fields such as immunology, materials science, and biology. In the future, advancing interdisciplinary collaboration among immunologists, materials scientists, and biologists is crucial to overcoming challenges and accelerating the clinical translation of nanomaterial-based cancer vaccines. Additionally, exploring synergistic combinations with other immunotherapeutic agents may enhance the efficacy of these vaccines in treating a broad spectrum of cancers.

Declarations

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

Parsa Ebrahimi Tirtashi led the conceptualization, while Seyedeh Ghazaleh Angaji, Mohammad Amin Salim, Alireza Azizi, and Negin Amiri contributed to the methodology. Saeede Rastakhiz, Negar Jahani, and Behnaz Akhlaghi conducted the formal analysis and investigation. The initial draft of the manuscript was prepared by Seyedeh Ghazaleh Angaji, Mohammad Amin Salim, Alireza Azizi, and Negin Amiri, with subsequent review and editing carried out by Saeede Rastakhiz, Negar Jahani, and Behnaz Akhlaghi. Parsa Ebrahimi Tirtashi provided supervision throughout the research process. All authors checked and approved the final version of the manuscript for publication in the present journal.

Funding

No funding was received for conducting this study.

Availability of data and materials

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethical considerations

The authors checked for plagiarism and consented to publish the article. The authors have also reviewed the article for data fabrication, double publication, and redundancy.

Acknowledgments

Not applicable.

References

- Farhood B, Geraily G, and Alizadeh A. Incidence and mortality of various cancers in Iran and compare to other countries: A review article. *Iran J Public Health*. 2018; 47(3): 309-316. PMID: <https://pubmed.ncbi.nlm.nih.gov/29845017>
- Qasemi A, Lagzian M, Bayat Z. Cancer and COVID-19: a double burden on the healthcare system. *Iran Red Crescent Med J*. 2023; 25(2): e2662. DOI: [10.32592/ircmj.2023.25.2.2662](https://doi.org/10.32592/ircmj.2023.25.2.2662)
- He J, Hu Y, Hu M, and Li B. Development of PD-1/PD-L1 pathway in tumor immune microenvironment and treatment for non-small cell lung cancer. *Sci Rep*. 2015; 5(1): 13110. DOI: [10.1038/srep13110](https://doi.org/10.1038/srep13110)
- Asouli A, Sadr S, Mohebalian H, and Borji H. Anti-Tumor Effect of Protoscolex Hydatid Cyst Somatic Antigen on Inhibition Cell Growth of K562. *Acta Parasitol*. 2023; 68: 385-392. DOI: [10.1007/s11686-023-00680-3](https://doi.org/10.1007/s11686-023-00680-3)
- Sadr S, Yousefsani Z, Simab PA, Alizadeh AJR, Lotfalizadeh N, and Borji H. *Trichinella spiralis* as a potential anti-tumor agent: An update. *World's Vet J*. 2023; 13(1): 65-74. DOI: [10.54203/scil.2023.wvj7](https://doi.org/10.54203/scil.2023.wvj7)
- Sadr S, Ghiassi S, Lotfalizadeh N, Simab PA, Hajjafari A, and Borji H. Anti-tumor mechanisms of molecules secreted by *Trypanosoma cruzi* in colon and breast cancer: A review. *Anti-Cancer Agents Med Chem*. 2023; 23(15): 1710-1721. DOI: [10.2174/1871520623666230529141544](https://doi.org/10.2174/1871520623666230529141544)
- Sadr S, and Borji H. *Echinococcus granulosus* as a promising therapeutic agent against triplenegative breast cancer. *Curr Cancer Ther Rev*. 2023; 19(4): 292-297. DOI: [10.2174/1573394719666230427094247](https://doi.org/10.2174/1573394719666230427094247)
- Sadr S, Poorjafari Jafroodi P, Haratizadeh MJ, Ghasemi Z, Borji H, and Hajjafari A. Current status of nano-vaccinology in veterinary medicine science. *Vet Med Sci*. 2023; 9(5): 2294-2308. DOI: [10.1002/vms3.1221](https://doi.org/10.1002/vms3.1221)
- Showalter A, Limaye A, Oyer JL, Igarashi R, Kittipatarin C, Copik AJ, et al. Cytokines in immunogenic cell death: Applications for cancer immunotherapy. *Cytokine*. 2017; 97: 123-132. DOI: [10.1016/j.cyto.2017.05.024](https://doi.org/10.1016/j.cyto.2017.05.024)
- Marshall J. Carcinoembryonic antigen-based vaccines. *Semin Oncol*. 2003; 30(3 Suppl 8): 30-36. DOI: [10.1016/S0093-7754\(03\)00233-1](https://doi.org/10.1016/S0093-7754(03)00233-1)
- Azmi F, Ahmad Fuaad AAH, Skwarczynski M, and Toth I. Recent progress in adjuvant discovery for peptide-based subunit vaccines. *Hum Vaccin Immunother*. 2014; 10(3): 778-796. DOI: [10.4161/hv.27332](https://doi.org/10.4161/hv.27332)
- Huang Y, Babiuk LA, and van Drunen Littel-van den Hurk S. The cell-mediated immune response induced by plasmid encoding bovine herpesvirus 1 glycoprotein B is enhanced by plasmid encoding IL-12 when delivered intramuscularly or by gene gun, but not after intradermal injection. *Vaccine*. 2006; 24(25): 5349-5359. DOI: [10.1016/j.vaccine.2006.04.026](https://doi.org/10.1016/j.vaccine.2006.04.026)
- Salvador A, Igartua M, Hernández RM, and Pedraz JL. An overview on the field of micro-and nanotechnologies for synthetic peptide-based vaccines. *J Drug Deliv*. 2011; 2011: 181646. DOI: [10.1155/2011/181646](https://doi.org/10.1155/2011/181646)
- Yang J, Firdaus F, Azuar A, Khalil ZG, Marasini N, Capon RJ, et al. Cell-penetrating peptides-based liposomal delivery system enhanced immunogenicity of peptide-based vaccine against Group A *Streptococcus*. *Vaccines*. 2021; 9(5): 499. DOI: [10.3390/vaccines9050499](https://doi.org/10.3390/vaccines9050499)
- Davies AM, Weinberg U, and Palti Y. Tumor treating fields: A new frontier in cancer therapy. *Ann N Y Acad Sci*. 2013; 1291(1): 86-95. DOI: [10.1111/nyas.12112](https://doi.org/10.1111/nyas.12112)
- Lindley C, McCune JS, Thomason TE, Lauder D, Sauls A, Adkins S, et al. Perception of chemotherapy side effects cancer versus noncancer patients. *Cancer Pract*. 1999; 7(2): 59-65. DOI: [10.1046/j.1523-5394.1999.07205.x](https://doi.org/10.1046/j.1523-5394.1999.07205.x)
- Ohnishi S, and Takeda H. Herbal medicines for the treatment of cancer chemotherapy-induced side effects. *Front Pharmacol*. 2015; 6: 14. DOI: [10.3389/fphar.2015.00014](https://doi.org/10.3389/fphar.2015.00014)
- Sohail M, Guo W, Li Z, Xu H, Zhao F, Chen D, et al. Nanocarrier-based drug delivery system for cancer therapeutics: A review of the last decade. *Curr Med Chem*. 2021; 28(19): 3753-3772. DOI: [10.2174/0929867327666201005111722](https://doi.org/10.2174/0929867327666201005111722)
- Sousa F, Ferreira D, Reis S, and Costa P. Current insights on antifungal therapy: Novel nanotechnology approaches for drug delivery systems and new drugs from natural sources. *Pharmaceuticals*. 2020; 13(9): 248. DOI: [10.3390/ph13090248](https://doi.org/10.3390/ph13090248)
- Saeed M, Sadr S, Gharib A, Lotfalizadeh N, Hajjafari A, Simab PA, et al. Phytosomes: A promising nanocarrier for enhanced delivery of herbal compounds in cancer therapy. *J Lab Anim Res*. 2022; 1(1): 26-32. DOI: [10.58803/jlar.v1i1.8](https://doi.org/10.58803/jlar.v1i1.8)
- Vela Ramirez JE, Roychoudhury R, Habte HH, Cho MW, Pohl NLB, and Narasimhan B. Carbohydrate-functionalized nanovaccines preserve HIV-1 antigen stability and activate antigen presenting cells. *J Biomater Sci Polym Edn*. 2014; 25(13): 1387-1406. DOI: [10.1080/09205063.2014.940243](https://doi.org/10.1080/09205063.2014.940243)
- Goradel NH, Nemati M, Bakhshandeh A, Arashkia A, and Negahdari B. Nanovaccines for cancer immunotherapy: Focusing on complex formation between adjuvant and antigen. *Int Immunopharmacol*. 2023; 117: 109887. DOI: [10.1016/j.intimp.2023.109887](https://doi.org/10.1016/j.intimp.2023.109887)
- Waldman AD, Fritz JM, and Lenardo MJ. A guide to cancer immunotherapy: From T cell basic science to clinical practice. *Nat Rev Immunol*. 2020; 20(11): 651-668. DOI: [10.1038/s41577-020-0306-5](https://doi.org/10.1038/s41577-020-0306-5)
- Linnemann C, van Buuren MM, Bies L, Verdegaal EME, Schotte R, Calis JJA, et al. High-throughput epitope discovery reveals frequent recognition of neo-antigens by CD4+ T cells in human melanoma. *Nat Med*. 2015; 21(1): 81-85. DOI: [10.1038/nm.3773](https://doi.org/10.1038/nm.3773)
- Gao J, Shi LZ, Zhao H, Chen J, Xiong L, He Q, et al. Loss of IFN-γ pathway genes in tumor cells as a mechanism of resistance to anti-CTLA-4 therapy. *Cell*. 2016; 167(2): 397-404. DOI: [10.1016/j.cell.2016.08.069](https://doi.org/10.1016/j.cell.2016.08.069)
- Li SD, Ma M, Li H, Waluszko A, Sidorenko T, Schadt EE, et al. Cancer gene profiling in non-small cell lung cancers reveals activating mutations in JAK2 and JAK3 with therapeutic implications. *Genome Med*. 2017; 9: 89. DOI: [10.1186/s13073-017-0478-1](https://doi.org/10.1186/s13073-017-0478-1)
- Chan TA, Yarchoan M, Jaffee E, Swanton C, Quezada SA, Stenzinger A, et al. Development of tumor mutation burden as an immunotherapy biomarker: Utility for the oncology clinic. *Ann Oncol*. 2019; 30(1): 44-56. DOI: [10.1093/annonc/mdy495](https://doi.org/10.1093/annonc/mdy495)
- Liu S, Lachapelle J, Leung S, Gao D, Foulkes WD, and Nielsen TO. CD8+ lymphocyte infiltration is an independent favorable prognostic indicator in basal-like breast cancer. *Breast Cancer Res*. 2012; 14: R48. DOI: [10.1186/bcr3148](https://doi.org/10.1186/bcr3148)
- Mahoney KM, Freeman GJ, and McDermott DF. The next immune-checkpoint inhibitors: PD-1/PD-L1 blockade in melanoma. *Clin Ther*. 2015; 37(4): 764-782. DOI: [10.1016/j.clinthera.2015.02.018](https://doi.org/10.1016/j.clinthera.2015.02.018)
- Festino L, Botti G, Lorigan P, Masucci GV, Hipp JD, Horak CE, et al. Cancer Treatment with Anti-PD-1/PD-L1 Agents: Is PD-L1 Expression a Biomarker for Patient Selection?. *Drugs*. 2016; 76(9): 925-945. DOI: [10.1007/s40265-016-0588-x](https://doi.org/10.1007/s40265-016-0588-x)
- Chow A, Perica K, Klebanoff CA, and Wolchok JD. Clinical implications of T cell exhaustion for cancer immunotherapy. *Nat Rev Clin Oncol*. 2022; 19(12): 775-790. DOI: [10.1038/s41571-022-00689-z](https://doi.org/10.1038/s41571-022-00689-z)
- Mahoney KM, Rennert PD, and Freeman GJ. Combination cancer immunotherapy and new immunomodulatory targets. *Nat Rev Drug Discov*. 2015; 14(8): 561-584. DOI: [10.1038/nrd4591](https://doi.org/10.1038/nrd4591)
- Farkona S, Diamandis EP, and Blasutig IM. Cancer immunotherapy: The beginning of the end of cancer?. *BMC Medicine*. 2016; 14(1): 73. DOI: [10.1186/s12916-016-0623-5](https://doi.org/10.1186/s12916-016-0623-5)
- Bloom DE, Canning D, and Weston M. The value of vaccination. Fighting the diseases of poverty. Routledge; 2017. p. 214-238. DOI: [10.4324/9780203791950-8](https://doi.org/10.4324/9780203791950-8)
- Lederberg J. Infectious history. *Science*. 2000; 288(5464): 287-293. DOI: [10.1126/science.288.5464.287](https://doi.org/10.1126/science.288.5464.287)
- Villain P, Gonzalez P, Almonte M, Franceschi S, Dillner J, Anttila A, et al. European code against cancer. *Cancer Epidemiol*. 2015; 39(S1): S120-S38. DOI: [10.1016/j.canep.2015.10.006](https://doi.org/10.1016/j.canep.2015.10.006)
- Disis ML. Mechanism of action of immunotherapy. *Semin Oncol*. 2014; 41(5): S3-S13. DOI: [10.1053/j.seminoncol.2014.09.004](https://doi.org/10.1053/j.seminoncol.2014.09.004)
- Palucka AK, Ueno H, Fay JW, and Banchereau J. Taming cancer by inducing immunity via dendritic cells. *Immunol Rev*. 2007; 220(1): 129-150. DOI: [10.1111/j.1600-065X.2007.00575.x](https://doi.org/10.1111/j.1600-065X.2007.00575.x)
- Borst J, Ahrends T, Bąbala N, Melief CJM, and Kastenmüller W. CD4+ T

- cell help in cancer immunology and immunotherapy. *Nat Rev Immunol*. 2018; 18(10): 635-647. DOI: [10.1038/s41577-018-0044-0](https://doi.org/10.1038/s41577-018-0044-0)
40. Faghfuri E, Pourfarzi F, Faghfouri AH, Abdoli Shadbad M, Hajiasgharzadeh K, and Baradaran B. Recent developments of RNA-based vaccines in cancer immunotherapy. *Expert Opin Biol Ther*. 2021; 21(2): 201-218. DOI: [10.1080/14712598.2020.1815704](https://doi.org/10.1080/14712598.2020.1815704)
41. Geng J, Xia X, Teng L, Wang L, Chen L, Guo X, et al. Emerging landscape of cell-penetrating peptide-mediated nucleic acid delivery and their utility in imaging, gene-editing, and RNA-sequencing. *J Control Release*. 2022; 341: 166-183. DOI: [10.1016/j.jconrel.2021.11.032](https://doi.org/10.1016/j.jconrel.2021.11.032)
42. Hobernik D, and Bros M. DNA vaccines—how far from clinical use?. *Int J Mol Sci*. 2018; 19(11) : 3605. DOI: [10.3390/ijms19113605](https://doi.org/10.3390/ijms19113605)
43. Trimble CL, Morrow MP, Kravnyak KA, Shen X, Dallas M, Yan J, et al. Safety, efficacy, and immunogenicity of VGX-3100, a therapeutic synthetic DNA vaccine targeting human papillomavirus 16 and 18 E6 and E7 proteins for cervical intraepithelial neoplasia 2/3: A randomised, double-blind, placebo-controlled phase 2b trial. *Lancet*. 2015; 386(10008): 2078-2088. DOI: [10.1016/S0140-6736\(15\)00239-1](https://doi.org/10.1016/S0140-6736(15)00239-1)
44. Kariko K, and Weissman D. Naturally occurring nucleoside modifications suppress the immunostimulatory activity of RNA: Implication for therapeutic RNA development. *Curr Opin Drug Discov Del*. 2007; 10(5): 523-532.
45. Hollingsworth RE, Jansen K. Turning the corner on therapeutic cancer vaccines. *npj Vaccines*. 2019; 4(1): 7. DOI: [10.1038/s41541-019-0103-y](https://doi.org/10.1038/s41541-019-0103-y)
46. Kübler H, Scheel B, Gnad-Vogt U, Miller K, Schultze-Seemann W, Vom Dorp F, et al. Self-adjuvanted mRNA vaccination in advanced prostate cancer patients: a first-in-man phase I/IIa study. *J Immunother Cancer*. 2015; 3(1): 1-14. DOI: [10.1186/s40425-015-0068-y](https://doi.org/10.1186/s40425-015-0068-y)
47. Bonilla WV, Kirchhammer N, Marx AF, Kallert SM, Krzyzaniak MA, Lu M, et al. Heterologous arenavirus vector prime-boost overrules self-tolerance for efficient tumor-specific CD8 T cell attack. *Cell Rep Med*. 2021; 2(3): 100209. DOI: [10.1016/j.xcrm.2021.100209](https://doi.org/10.1016/j.xcrm.2021.100209)
48. Zom GG, Khan S, Filippov DV, and Ossendorp F. TLR ligand-peptide conjugate vaccines: toward clinical application. *Adv Immunol*. 2012; 114: 177-201. DOI: [10.1016/B978-0-12-396548-6.00007-X](https://doi.org/10.1016/B978-0-12-396548-6.00007-X)
49. Dorostkar F, Arashkia A, Roohvand F, Shoja Z, Navari M, Mashhadi Abolghasem Shirazi M, et al. Co-administration of 2'3'-cGAMP STING activator and CpG-C adjuvants with a mutated form of HPV 16 E7 protein leads to tumor growth inhibition in the mouse model. *Infectious Agents and Cancer*. 2021; 16(1): 7. Available at: <https://infectagentscancer.biomedcentral.com/articles/10.1186/s13027-021-00346-7>
50. Rosalia RA, Quakkelaar ED, Redeker A, Khan S, Camps M, Drijfhout JW, et al. Dendritic cells process synthetic long peptides better than whole protein, improving antigen presentation and T-cell activation. *Eur J Immunol*. 2013; 43(10): 2554-2565. DOI: [10.1002/eji.201343324](https://doi.org/10.1002/eji.201343324)
51. Blass E, and Ott PA. Advances in the development of personalized neoantigen-based therapeutic cancer vaccines. *Nat Rev Clin Oncol*. 2021; 18(4): 215-229. DOI: [10.1038/s41571-020-00460-2](https://doi.org/10.1038/s41571-020-00460-2)
52. Demento SL, Cui W, Criscione JM, Stern E, Tulipan J, Kaech SM, et al. Role of sustained antigen release from nanoparticle vaccines in shaping the T cell memory phenotype. *Biomaterials*. 2012; 33(19): 4957-4964. DOI: [10.1016/j.biomaterials.2012.03.041](https://doi.org/10.1016/j.biomaterials.2012.03.041)
53. Dhakal S, and Renukaradhya GJ. Nanoparticle-based vaccine development and evaluation against viral infections in pigs. *Vet Res*. 2019; 50(1): 90. DOI: [10.1186/s13567-019-0712-5](https://doi.org/10.1186/s13567-019-0712-5)
54. Cha BG, Jeong JH, and Kim J. Extra-large pore mesoporous silica nanoparticles enabling co-delivery of high amounts of protein antigen and toll-like receptor 9 agonist for enhanced cancer vaccine efficacy. *ACS Cent Sci*. 2018; 4(4): 484-492. DOI: [10.1021/acscentsci.8b00035](https://doi.org/10.1021/acscentsci.8b00035)
55. Hong X, Zhong X, Du G, Hou Y, Zhang Y, Zhang Z, et al. The pore size of mesoporous silica nanoparticles regulates their antigen delivery efficiency. *Sci Adv*. 2020; 6(25): eaaz4462. DOI: [10.1126/sciadv.aaz4462](https://doi.org/10.1126/sciadv.aaz4462)
56. Xiang SD, Scholzen A, Minigo G, David C, Apostolopoulos V, Mottram PL, et al. Pathogen recognition and development of particulate vaccines: does size matter?. *Methods*. 2006; 40(1): 1-9. DOI: [10.1016/j.ymeth.2006.05.016](https://doi.org/10.1016/j.ymeth.2006.05.016)
57. Danaei M, Dehghankhold M, Ataei S, Hasanizadeh Davarani F, Javanmard R, Dokhani A, et al. Impact of particle size and polydispersity index on the clinical applications of lipidic nanocarrier systems. *Pharmaceutics*. 2018; 10(2): 57. DOI: [10.3390/pharmaceutics10020057](https://doi.org/10.3390/pharmaceutics10020057)
58. Sarin H, Kanevsky AS, Wu H, Sousa AA, Wilson CM, Aronova MA, et al. Physiologic upper limit of pore size in the blood-tumor barrier of malignant solid tumors. *J Transl Med*. 2009; 7(1): 51. DOI: [10.1186/1479-2875-7-51](https://doi.org/10.1186/1479-2875-7-51)
59. He C, Hu Y, Yin L, Tang C, and Yin C. Effects of particle size and surface charge on cellular uptake and biodistribution of polymeric nanoparticles. *Biomaterials*. 2010; 31(13): 3657-3666. DOI: [10.1016/j.biomaterials.2010.01.065](https://doi.org/10.1016/j.biomaterials.2010.01.065)
60. Silva JM, Vandermeulen G, Oliveira VG, Pinto SN, Rodrigues C, Salgado A, et al. Development of functionalized nanoparticles for vaccine delivery to dendritic cells: a mechanistic approach. *Nanomedicine*. 2014; 9(17): 2639-2656. DOI: [10.2217/nmm.14.135](https://doi.org/10.2217/nmm.14.135)
61. Zaman M, Good MF, and Toth I. Nanovaccines and their mode of action. *Methods*. 2013; 60(3): 226-231. DOI: [10.1016/j.ymeth.2013.04.014](https://doi.org/10.1016/j.ymeth.2013.04.014)
62. Hwang I, Huang J-F, Kishimoto H, Brunmark A, Peterson PA, Jackson MR, et al. T cells can use either T cell receptor or CD28 receptors to absorb and internalize cell surface molecules derived from antigen-presenting cells. *J Exp Med*. 2000; 191(7): 1137-1148. DOI: [10.1084/jem.191.7.1137](https://doi.org/10.1084/jem.191.7.1137)
63. Nooraei S, Sarkar Lotfabadi A, Akbarzadehmoallemkolaei M, and Rezaei N. Immunogenicity of different types of adjuvants and nano-adjuvants in veterinary vaccines: A comprehensive review. *Vaccines*. 2023; 11(2): 453. DOI: [10.3390/vaccines11020453](https://doi.org/10.3390/vaccines11020453)
64. Ilyinski PO, Roy CJ, O'Neil CP, Browning EA, Pittet LA, Altmutter DH, et al. Adjuvant-carrying synthetic vaccine particles augment the immune response to encapsulated antigen and exhibit strong local immune activation without inducing systemic cytokine release. *Vaccine*. 2014; 32(24): 2882-2895. DOI: [10.1016/j.vaccine.2014.02.027](https://doi.org/10.1016/j.vaccine.2014.02.027)
65. Joffe O, Nolte MA, Spörri R, and Sousa CRE. Inflammatory signals in dendritic cell activation and the induction of adaptive immunity. *Immunol Rev*. 2009; 227(1): 234-247. DOI: [10.1111/j.1600-065X.2008.00718.x](https://doi.org/10.1111/j.1600-065X.2008.00718.x)
66. Harper J, Sainson RC, editors. Regulation of the anti-tumour immune response by cancer-associated fibroblasts. *Semin Cancer Biol*. 2014; 25: 69-77. DOI: [10.1016/j.semcancer.2013.12.005](https://doi.org/10.1016/j.semcancer.2013.12.005)
67. Zamani P, Navashenaq JG, Teymouri M, Karimi M, Mashreghi M, and Jaafari MR. Combination therapy with liposomal doxorubicin and liposomal vaccine containing E75, an HER-2/neu-derived peptide, reduces myeloid-derived suppressor cells and improved tumor therapy. *Life Sci*. 2020; 252: 117646. DOI: [10.1016/j.lfs.2020.117646](https://doi.org/10.1016/j.lfs.2020.117646)
68. Chu Y, Qian L, Ke Y, Feng X, Chen X, Liu F, et al. Lymph node-targeted neoantigen nanovaccines potentiate anti-tumor immune responses of post-surgical melanoma. *J Nanobiotechnology*. 2022; 20(1): 190. DOI: [10.1186/s12951-022-01397-7](https://doi.org/10.1186/s12951-022-01397-7)
69. Danielsson R, and Eriksson H. Aluminium adjuvants in vaccines—A way to modulate the immune response. *Semin Cell Dev Biol*. 2021; 115: 3-9. DOI: [10.1016/j.semcdb.2020.12.008](https://doi.org/10.1016/j.semcdb.2020.12.008)
70. Vecchiarelli A. Cytokines and costimulatory molecules: Positive and negative regulation of the immune response to *Cryptococcus neoformans*. In: Górski A, Krotkiewski H, Zimecki M, editors. *Inflammation*. Dordrecht: Springer; 2001. p. 51-65. DOI: [10.1007/978-94-015-9702-9_5](https://doi.org/10.1007/978-94-015-9702-9_5)
71. Gavas S, Quazi S, and Karpiński TM. Nanoparticles for cancer therapy: Current progress and challenges. *Nanoscale Res Lett*. 2021; 16(1): 173. DOI: [10.1186/s11671-021-03628-6](https://doi.org/10.1186/s11671-021-03628-6)
72. Liu J, Fu M, Wang M, Wan D, Wei Y, and Wei X. Cancer vaccines as promising immuno-therapeutics: Platforms and current progress. *J Hematol Oncol*. 2022; 15(1): 28. DOI: [10.1186/s13045-022-01247-x](https://doi.org/10.1186/s13045-022-01247-x)
73. Cui X, and Snapper CM. Epstein Barr virus: Development of vaccines and immune cell therapy for EBV-associated diseases. *Front Immunol*. 2021; 12: 734471. DOI: [10.3389/fimmu.2021.734471](https://doi.org/10.3389/fimmu.2021.734471)
74. Mallilankaraman K, Shedlock DJ, Bao H, Kawalekar OU, Fagone P, Ramanathan AA, et al. A DNA vaccine against chikungunya virus is protective in mice and induces neutralizing antibodies in mice and nonhuman primates. *PLoS Negl Trop Dis*. 2011; 5(1): e928. DOI: [10.1371/journal.pntd.0000928](https://doi.org/10.1371/journal.pntd.0000928)
75. Zhong L, Krummenacher C, Zhang W, Hong J, Feng Q, Chen Y, et al. Urgency and necessity of Epstein-Barr virus prophylactic vaccines. *npj Vaccines*. 2022; 7(1): 159. DOI: [10.1038/s41541-022-00587-6](https://doi.org/10.1038/s41541-022-00587-6)
76. Parvanian S, Mostafavi SM, and Aghashiri M. Multifunctional nanoparticle developments in cancer diagnosis and treatment. *Sens Bio-Sens Res*. 2017; 13: 81-87. DOI: [10.1016/j.sbsr.2016.08.002](https://doi.org/10.1016/j.sbsr.2016.08.002)
77. Nicholaou T, Ebert L, Davis ID, Robson N, Klein O, Maraskovsky E, et al. Directions in the immune targeting of cancer: Lessons learned from the cancer-testis Ag NY-ESO-1. *Immunol Cell Biol*. 2006; 84(3): 303-317. DOI: [10.1111/j.1440-1711.2006.01446.x](https://doi.org/10.1111/j.1440-1711.2006.01446.x)
78. Koerner J, Horvath D, Herrmann VL, MacKerracher A, Gander B, Yagita H, et al. PLGA-particle vaccine carrying TLR3/RIG-I ligand Riboxim

- synergizes with immune checkpoint blockade for effective anti-cancer immunotherapy. *Nat Commun.* 2021; 12(1): 2935. DOI: [10.1038/s41467-021-23244-3](https://doi.org/10.1038/s41467-021-23244-3)
79. Neshat SY, Tzeng SY, and Green JJ. Gene delivery for immunoengineering. *Curr Opin Biotechnol.* 2020; 66: 1-10. DOI: [10.1016/j.copbio.2020.05.008](https://doi.org/10.1016/j.copbio.2020.05.008)
80. Blanco E, Chocarro L, Fernández-Rubio L, Bocanegra A, Arasanz H, Echaide M, et al. Leading edge: Intratumor delivery of monoclonal antibodies for the treatment of solid tumors. *Int J Mol Sci.* 2023; 24(3): 2676. DOI: [10.3390/ijms24032676](https://doi.org/10.3390/ijms24032676)
81. Alberer M, Gnad-Vogt U, Hong HS, Mehr KT, Backert L, Finak G, et al. Safety and immunogenicity of a mRNA rabies vaccine in healthy adults: An open-label, non-randomised, prospective, first-in-human phase 1 clinical trial. *Lancet.* 2017; 390(10101): 1511-1520. DOI: [10.1016/S0140-6736\(17\)31665-3](https://doi.org/10.1016/S0140-6736(17)31665-3)
82. Pound P, Ritskes-Hoitinga M. Is it possible to overcome issues of external validity in preclinical animal research? Why most animal models are bound to fail. *J Transl Med.* 2018; 16(1): 304. DOI: [10.1186/s12967-018-1678-1](https://doi.org/10.1186/s12967-018-1678-1)
83. Aikins ME, Xu C, and Moon JJ. Engineered nanoparticles for cancer vaccination and immunotherapy. *Acc Chem Resh.* 2020; 53(10): 2094-2105. DOI: [10.1021/acs.accounts.0c00456](https://doi.org/10.1021/acs.accounts.0c00456)
84. Gao S, Yang D, Fang Y, Lin X, Jin X, Wang Q, et al. Engineering nanoparticles for targeted remodeling of the tumor microenvironment to improve cancer immunotherapy. *Theranostics.* 2019; 9(1): 126-151. Available at: <https://www.thno.org/v09p0126.htm>
85. Goldblatt EM, and Lee W-H. From bench to bedside: The growing use of translational research in cancer medicine. *Am J Transl Res.* 2010; 2(1):1-8. PMID: <https://pubmed.ncbi.nlm.nih.gov/20182579>
86. Denayer T, Stöhr T, and Van Roy M. Animal models in translational medicine: Validation and prediction. *New Horizons in Translational Medicine.* 2014; 2(1): 5-11. DOI: [10.1016/j.nhtm.2014.08.001](https://doi.org/10.1016/j.nhtm.2014.08.001)
87. Zhang Z, Lu M, Qin Y, Gao W, Tao L, Su W, et al. Neoantigen: A new breakthrough in tumor immunotherapy. *Front Immunol.* 2021; 12: 672356. DOI: [10.3389/fimmu.2021.672356](https://doi.org/10.3389/fimmu.2021.672356)
88. Vignais PV, and Vignais PM. Challenges for experimentation on living beings at the dawn of the 21 st century. *Discovering Life, Manufacturing Life.* Dordrecht: Springer; 2010. p. 241-345. DOI: [10.1007/978-90-481-3767-1_5](https://doi.org/10.1007/978-90-481-3767-1_5)
89. Luo M, Feng Y, Wang T, and Guan J. Micro-/nanorobots at work in active drug delivery. *Adv Funct Mater.* 2018; 28(25): 1706100. DOI: [10.1002/adfm.201706100](https://doi.org/10.1002/adfm.201706100)